



Are neonicotinoids a threat for leaf litter breakdown? A laboratory approach with leaf shredding invertebrates

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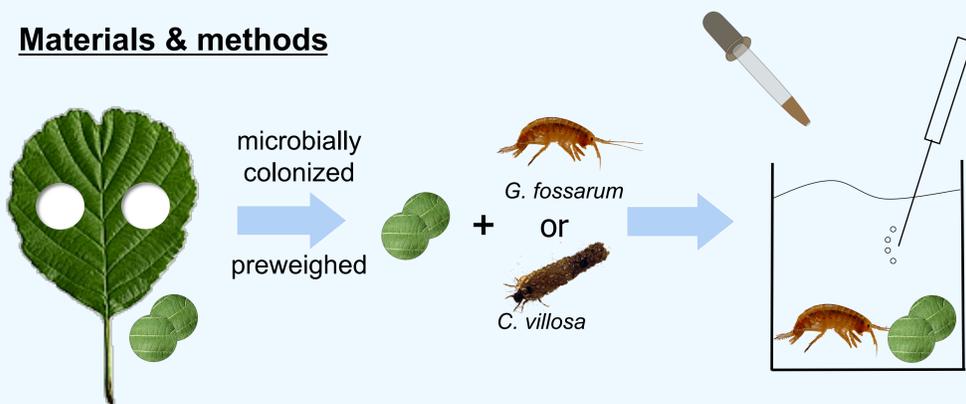
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Introduction & objectives

Neonicotinoids are registered in over 120 countries for use against herbivorous insects in agriculture, horticulture, forestry, and tree nursery [1]. Due to their extensive application and chemical properties, neonicotinoids are susceptible to be transported into surface waters via various pathways [2]. There they might pose a threat to non-target species and associated ecosystem functions, such as the breakdown of leaf litter. This particular ecosystem function is critical in providing energy for local as well as downstream communities [3]. A) To assess for potential risks of three frequently used neonicotinoid insecticides (i.e. thiacloprid (THI), imidacloprid (IMI) and acetamiprid (ACE)) in this process, the feeding rate of *Gammarus fossarum* (Amphipoda) and *Chaetopteryx villosa* (Trichoptera) – two key leaf-shredding macroinvertebrates [4] – was assessed over seven days. B) As neonicotinoids are regularly detected as mixtures in the aquatic environment, an additional experiment accounted for the joint toxicity of a mixture composed of THI, IMI and ACE on the feeding rate of *G. fossarum*.

Materials & methods



A) single substance experiments with *G. fossarum* and *C. villosa*

- 3 neonicotinoids (THI, IMI, ACE)
- 6 concentrations (between 0-24 µg/L; n=30)
- endpoint: feeding inhibition (7d-EC_{20,50})

B) mixture experiment with *G. fossarum*

- similar experimental set up as used in A)
- mixture of THI, IMI, ACE based on EC₅₀s from A) [5]
- comparison with predictions by concentration addition model [6]

Tab. 1 Calculated 7d-EC₂₀ and EC₅₀ values (in µg/L) for *G. fossarum* and *C. villosa* after exposure to THI, IMI and ACE as well as the maximum detected field concentrations.

	<i>G. fossarum</i>		<i>C. villosa</i>		max. detected field concentrations
	EC ₂₀ ±95%CI	EC ₅₀ ±95%CI	EC ₂₀ ±95%CI	EC ₅₀ ±95%CI	
THI	1.7± 0.5	3.1± 0.8	4.5± 22.2		4.5 [7]
IMI	3.6± 1.7	8.3± 2.7	10.4± 2.3	19.3± 2.7	320 [8]
ACE	2.3± 2.2	8.4± 4.9	11.7± 11.1		44.1 [9]

Results & discussion

In all experiments exposure to neonicotinoids resulted in a concentration dependent decrease in the feeding rate of both shredders, while the amphipod *G. fossarum* was 3 to 5 times more sensitive (based on EC₂₀s) towards neonicotinoids than the caddisfly larvae *C. villosa* (Tab. 1). When comparing EC₂₀s, thiacloprid was – irrespective of the species investigated – the most toxic neonicotinoid. Moreover, the measured feeding rate of *G. fossarum* exposed to the neonicotinoid mixture agreed well with the predictions by the concentration addition model (i.e. prediction is within the 95% confidence intervals (CIs) of the observations – except for the highest concentration; Fig. 1).

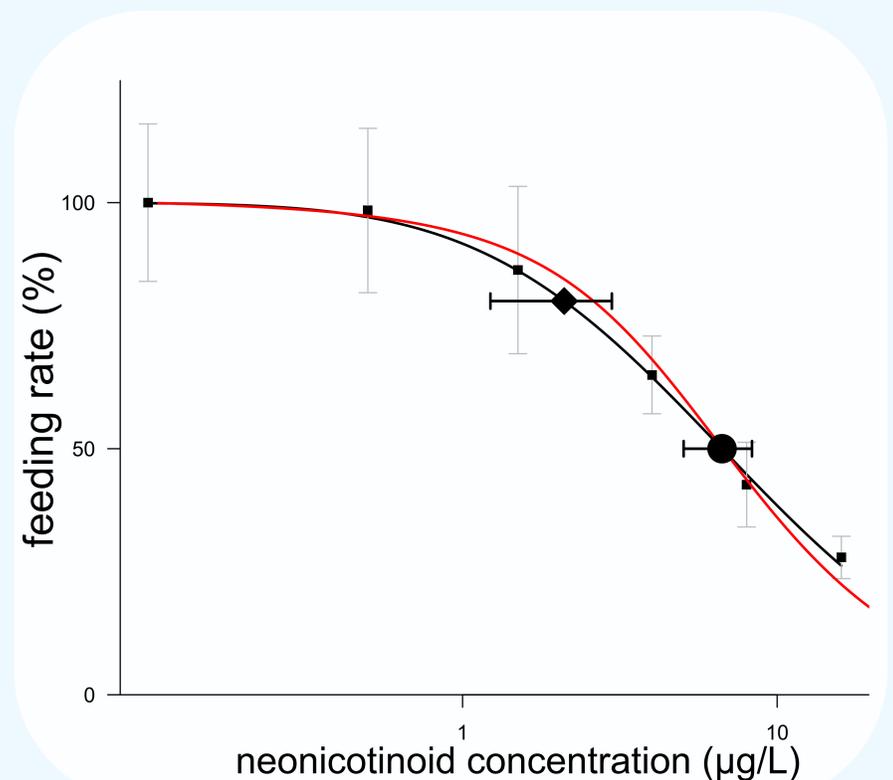


Fig. 1 Mean feeding rate of *G. fossarum* (■; ±95%CIs) exposed to the neonicotinoid mixture, the best fitting model (black line) and the predictions derived from the concentration addition model (red line). 7d-EC₂₀s (♦) and EC₅₀s (•) ±95%CIs are also displayed.

Conclusions

The present study indicates a risk of neonicotinoids and their mixtures at field relevant concentrations (Tab.1) to adversely affect leaf shredding invertebrates that are considered as key species in the ecosystem function of leaf litter breakdown. Such implications might restrict the energy availability (in the form of feces) e.g. for collectors and consequently reduce the prey availability (in the form of both shredders and collectors) for aquatic (e.g. fish [10]) and – following the emergence of insects (such as *C. villosa*) – terrestrial predators (e.g. spiders, birds, bats [11]).

References

- [1] Jeschke P, Nauen R. 2008. *Pest management science* 64: 1084-1098
- [2] Morrissey CA, Mineau P, Devries JH, Sanchez-Bayo F, Liess M, et al. 2015. *Environment international* 74C: 291-303
- [3] Cummins KW, Klug MJ. 1979. *Annual Review of Ecology, Evolution and Systematics* 10: 147-172
- [4] Dangles O, Malmqvist B. 2004. *Ecology letters* 7: 395-402
- [5] Jonker MJ, Backhaus GA, van Gestel CA. 2011. In *Mixture Toxicology*, ed. CA van Gestel, pp. 121-156. Boca Raton: CRC Press
- [6] Loewe S, Muischnek H. 1926. *Archiv für Experimentelle Pathologie und Pharmakologie* 114: 313-326
- [7] Süß A, Bischoff G, Mueller ACW, Buhr L. 2006. *Nachrichtenblatt des Deutschen Pflanzenschutzdienstes* 58: 28-42
- [8] Van Dijk TC, Van Staalduinen MA, Van der Sluijs JP. 2013. *PLoS One* 8
- [9] Anderson TA, Salice CJ, Erickson RA, McMurry ST, Cox SB, Smith LM. 2013. *Chemosphere* 92: 84-90
- [10] Wallace JB, Eggert SL, Meyer JL, Webster JR. 1997. *Science* 277: 102-104
- [11] Nakano S, Murakami M. 2001. *Proceedings of the National Academy of Sciences of the United States of America* 98: 166-170